LETTER

doi:10.1038/nature24659

A semi-synthetic organism that stores and retrieves increased genetic information

Yorke Zhang[1], Jerod L. Ptacin[2], Emil C. Fischer[1], Hans R. Aerni[2], Carolina E. Caffaro[2], Kristine San Jose[2], Aaron W. Feldman[1], Court R. Turner[2] & Floyd E. Romesberg[1]

Since at least the last common ancestor of all life on Earth, genetic information has been stored in a four-letter alphabet that is propagated and retrieved by the formation of two base pairs. The central goal of synthetic biology is to create new life forms and functions[1], and the most general route to this goal is the creation of semi-synthetic organisms whose DNA harbours two additional letters that form a third, unnatural base pair. Previous efforts to generate such semi-synthetic organisms[2] culminated in the creation of a strain of *Escherichia coli* that, by virtue of a nucleoside triphosphate transporter from *Phaeodactylum tricornutum*, imports the requisite unnatural triphosphates from its medium and then uses them to replicate a plasmid containing the unnatural base pair dNaM–dTPT3. Although the semi-synthetic organism stores increased information when compared to natural organisms, retrieval of the information requires *in vivo* transcription of the unnatural base pair into mRNA and tRNA, aminoacylation of the tRNA with a non-canonical amino acid, and efficient participation of the unnatural base pair in decoding at the ribosome. Here we report the *in vivo* transcription of DNA containing dNaM and dTPT3 into mRNAs with two different unnatural codons and tRNAs with cognate unnatural anticodons, and their efficient decoding at the ribosome to direct the site-specific incorporation of natural or non-canonical amino acids into superfolder green fluorescent protein. The results demonstrate that interactions other than hydrogen bonding can contribute to every step of information storage and retrieval. The resulting semi-synthetic organism both encodes and retrieves increased information and should serve as a platform for the creation of new life forms and functions.

Green fluorescent protein and variants such as superfolder green fluorescent protein (sfGFP)[3] have served as model systems for the study of the incorporation of non-canonical amino acids (ncAAs) via genetic code expansion[4], including at position Y151, which has been shown to tolerate a variety of natural amino acids and ncAAs[5] (Extended Data Fig. 1). To investigate the decoding of unnatural codons, we first focused on the incorporation of Ser at position 151 of sfGFP, as *E. coli* seryl-tRNA synthetase (SerRS) does not rely on anticodon recognition for tRNA aminoacylation[6], thus eliminating the potential complications of inefficient charging. The semi-synthetic organism (SSO) strain YZ3[2], carrying a plasmid expressing the Tet repressor protein TetR, was transformed with a plasmid encoding sfGFP and an *E. coli* $tRNA^{Ser}$ gene (*serT*), with the native sfGFP codon 151 (TAC) replaced by the unnatural codon AXC (sfGFP(AXC)^{151}; X denotes NaM), and the anticodon of *serT* replaced by the unnatural anticodon GYT ($tRNA^{Ser}$(GYT); Y denotes TPT3) (Fig. 1a, b). Transformants were grown in medium supplemented with dNaMTP and dTPT3TP, then supplemented further with NaMTP and TPT3TP, as well as isopropyl-β-D-thiogalactoside (IPTG) to induce expression of T7 RNA polymerase (T7 RNAP) and $tRNA^{Ser}$(GYT). After a brief period of tRNA induction, anhydrotetracycline (aTc) was added to induce expression of sfGFP(AXC)^{151}, and the production of sfGFP was monitored by fluorescence.

Following induction, cells transformed with a control plasmid encoding sfGFP(AXC)^{151} but lacking $tRNA^{Ser}$(GYT) showed greatly reduced fluorescence compared to cells transformed with a plasmid encoding sfGFP with a natural Ser codon at position 151 (sfGFP(AGT)^{151}; Fig. 1c). Moreover, cell growth began to plateau upon induction of sfGFP(AXC)^{151} (Fig. 1d), probably owing to the stalling and sequestering of ribosomes[7]; lysates of these cells subjected to western blotting with an anti-GFP antibody revealed a substantial reduction in sfGFP expression and the presence of sfGFP as truncated at the position of the unnatural codon (Fig. 1e). By contrast, cells transformed with the plasmid encoding both sfGFP(AXC)^{151} and $tRNA^{Ser}$(GYT) exhibited fluorescence that was nearly equal to that of control cells expressing sfGFP(AGT)^{151} (Fig. 1c), cell growth did not plateau upon induction of sfGFP(AXC)^{151} (Fig. 1d), and western blots of lysates from these cells revealed only full-length sfGFP protein (Fig. 1e). Furthermore, we assessed the ability of all four natural near-cognate tRNAs ($tRNA^{Ser}$(GNT); N denotes G, C, A or T), expressed in an identical fashion, to decode the AXC codon. In each case, low fluorescence was observed and the growth defect remained (Extended Data Fig. 2a, b). These data indicate that the nucleoside triphosphate transporter from *P. tricornutum* (*Pt*NTT2) is able to import both the deoxy- and ribotriphosphates of both unnatural nucleotides, that T7 RNAP is able to transcribe mRNA and tRNA containing the unnatural nucleotides *in vivo*, and that the ribosome efficiently decodes the unnatural codon only with an unnatural anticodon.

To assess the fidelity of decoding, we analysed protein purified from cells expressing both sfGFP(AXC)^{151} and $tRNA^{Ser}$(GYT) using liquid chromatography–tandem mass spectrometry (LC–MS/MS) and relative quantification via ion intensities. The results revealed 98.5 ± 0.7% (95% confidence interval (CI), $n = 4$) incorporation of Ser at position 151, with Ile/Leu being the predominant contaminant (Fig. 1f, Extended Data Table 1). Given that the retention of the unnatural base pair (UBP) in the sfGFP(AXC)^{151} gene was 98 ± 2% (95% CI, $n = 4$) (Extended Data Table 2) and that X→T is typically the major mutation during replication[2] (which for AXC would result in the Ile codon ATC), we attribute the majority of the protein not containing Ser at position 151 to loss of the UBP during replication and conclude that the fidelity of translation with the unnatural codon is high.

To demonstrate the encoding of ncAAs with UBPs, we constructed plasmids analogous to those used above, but with the $tRNA^{Ser}$ gene replaced with the *Methanosarcina mazei* $tRNA^{Pyl}$(GYT) gene. $tRNA^{Pyl}$ can be selectively charged by the *Methanosarcina barkeri* pyrrolysyl-tRNA synthetase (PylRS) with the ncAA N^6-[(2-propynyloxy)carbonyl]-L-lysine (PrK)[8,9]. In addition to the codon AXC, we also analysed the codon GXC and the corresponding $tRNA^{Pyl}$(GYC). The SSO, carrying a separate plasmid encoding an IPTG-inducible PylRS (expressed from a P_{tac} promoter), was transformed with the required plasmids and grown with or without added PrK. In control experiments with cells expressing either sfGFP(AXC)^{151} or sfGFP(GXC)^{151} in the absence of PylRS, the cognate unnatural $tRNA^{Pyl}$, or PrK, we observed only low

[1]Department of Chemistry, The Scripps Research Institute, La Jolla, California 92037, USA. [2]Synthorx, Inc., La Jolla, California 92037, USA.

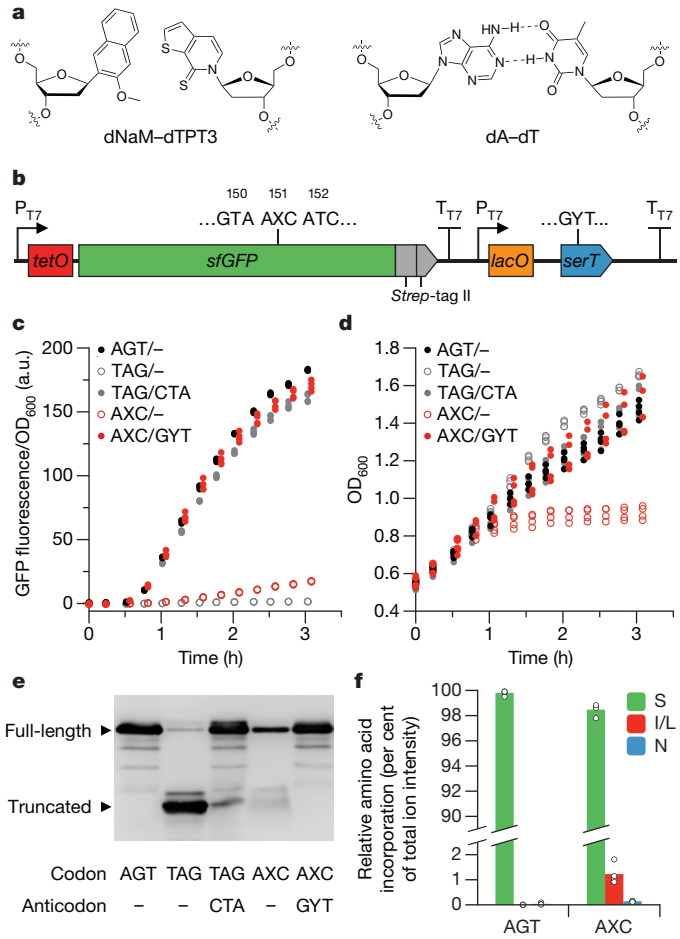


Figure 1 | UBP and decoding with tRNA^{Ser}. a, Chemical structure of the dNaM–dTPT3 UBP and a natural dA–dT base pair. **b**, Schematic illustration of the gene cassette used to express sfGFP(AXC)^{151} and tRNA^{Ser}(GYT). P_{T7} and T_{T7} denote the T7 RNAP promoter and terminator, respectively. In controls where sfGFP is expressed in the absence of *serT*, the sequence following the sfGFP T7 terminator is absent. **c**, **d**, Fluorescence (**c**) and growth (**d**) of cells expressing sfGFP and tRNA^{Ser} with the indicated position 151 codon and anticodon, respectively. Minus sign denotes the absence of *serT* in the expression cassette. $t = 0$ corresponds to the addition of IPTG to induce expression of T7 RNAP and tRNA^{Ser} (if present); aTc was added at $t = 0.5$ h to induce expression of sfGFP. Individual data points shown from $n = 4$ cultures, each propagated from an individual colony (biological replicates). a.u., arbitrary unit. **e**, Western blot of lysates (normalized by OD_{600}) from cells collected at the last time point shown in **c** and **d**, probed with an anti-GFP antibody (N-terminal epitope). For blot source data, see Supplementary Fig. 1. **f**, Relative abundance of amino acids (S, I/L and N) at position 151 of sfGFP purified from cells expressing sfGFP(AGT)^{151} or sfGFP(AXC)^{151} and tRNA^{Ser}(GYT), as determined by LC–MS/MS and precursor ion intensity-based quantification; amino acids detected at <0.1% (on average, for both codons) are not shown. Data shown as mean with individual values, $n = 4$ purified sfGFP samples, each from a culture propagated from an individual colony and collected at the last time point shown in **c** and **d**.

cellular fluorescence (Fig. 2a), truncation of sfGFP (Extended Data Fig. 3a), and a plateau in cell growth (Extended Data Fig. 3b). By contrast, for either unnatural mRNA with its cognate unnatural tRNA, when PylRS was present and PrK was added, we observed high fluorescence (64% and 69% of sfGFP(TAC)^{151} for AXC and GXC, respectively) (Fig. 2a, b), robust production of full-length sfGFP (Extended Data Fig. 3a), and normal growth (Extended Data Fig. 3b).

To verify the incorporation of PrK, sfGFP was affinity-purified from cell lysates using a tandem C-terminal *Strep*-tag II[10] and subjected to copper-catalysed click chemistry[11] to attach a carboxytetramethylrhodamine (TAMRA) dye (TAMRA-PEG_4-N_3), which was found to shift the electrophoretic mobility of sfGFP during SDS–PAGE, thus allowing us to assess the fidelity of PrK incorporation by western blotting (Fig. 2c). We observed a strong TAMRA signal and a shift of virtually all of the sfGFP when the protein was purified from cells that expressed sfGFP(AXC)^{151} and tRNA^{Pyl}(GYT) or sfGFP(GXC)^{151} and tRNA^{Pyl}(GYC), and that had been cultured in medium supplemented with PrK (Fig. 2c). By contrast, we observed little or no TAMRA signal or shifted sfGFP when NaMTP, TPT3TP, or both were absent (Extended Data Fig. 4a). Finally, no TAMRA signal nor shifted sfGFP was observed in protein purified from cells expressing sfGFP(TAC)^{151} with either unnatural tRNA (Fig. 2c). These data demonstrate that PrK is specifically incorporated into sfGFP through decoding of the unnatural codons by tRNAs with an unnatural anticodon.

With optimal PrK concentrations[12] (Extended Data Fig. 5), we purified 54 ± 4 and 55 ± 6 μg of sfGFP per millilitre of culture for the AXC and GXC codons, respectively (mean ± s.d., $n = 4$; approximately 40% of the sfGFP(TAC)^{151} control, Extended Data Table 3). Moreover, according to mass spectrometry analysis, the purity of sfGFP with PrK was 96.2 ± 0.3% (95% CI, $n = 4$) for the AXC codon and 97.5 ± 0.7% (95% CI, $n = 4$) for the GXC codon (Fig. 2d). As with the Ser incorporation experiments, the main natural amino acid contaminant corresponded to an X→T mutation (ATC, Ile; GTC, Val); however, we also observed an increase in the incorporation of amino acids that appeared to result from mischarging (Lys) or from mis-decoding by near-cognate natural tRNAs containing a pyrimidine at the second position (Extended Data Fig. 2; Fig. 2d), which is likely to result from inefficient charging of the heterologous tRNA or its reduced ability to compete with endogenous tRNAs[13]. Nonetheless, although the yield of sfGFP protein purified was slightly lower than that obtained with amber suppression (87 ± 6 μg ml^{-1} of culture; mean ± s.d., $n = 4$, Extended Data Table 3) owing to a moderate reduction in growth with the addition of the unnatural ribotriphosphates (Extended Data Fig. 4b, c), decoding of both unnatural codons resulted in higher fluorescence than observed with amber suppression when normalized to cell density (Fig. 2a, b), and less truncated protein (Extended Data Fig. 3a), implying that decoding with the unnatural codons is more efficient than amber suppression.

To explore the encoding of other ncAAs with UBPs, we examined the encoding of *p*-azido-phenylalanine (*p*AzF) with the AXC codon and an evolved *Methanococcus jannaschii* TyrRS–tRNA^{Tyr} pair[14] (*p*AzFRS–tRNA^{pAzF}). With induction of the synthetase (expressed from an IPTG-inducible P_{lacUV5} promoter) and the addition of *p*AzF to the growth medium, we observed robust fluorescence equivalent to that of cells expressing natural sfGFP(TAC)^{151} and normal growth with sfGFP(AXC)^{151} and tRNA^{pAzF}(GYT) (Fig. 3a; Extended Data Fig. 6). Full-length sfGFP was purified (86 ± 6 μg ml^{-1} of culture; mean ± s.d., $n = 4$; 68% of the sfGFP(TAC)^{151} control, Extended Data Table 3) and subjected to copper-free click chemistry using a dibenzocyclooctyl (DBCO) group to attach TAMRA (TAMRA-PEG_4-DBCO)[15]. We observed robust TAMRA conjugation to sfGFP isolated from cells expressing sfGFP(AXC)^{151} and tRNA^{pAzF}(GYT) and cultured in the presence of *p*AzF (Fig. 3b). Although we were unable to accurately assess the fidelity of *p*AzF incorporation, owing to decomposition of the azido moiety[16,17], about 93% of the sfGFP protein observed in the western blot was shifted, which compares favourably to the approximately 95% shifted sfGFP produced via amber suppression (Fig. 3b).

Since at least the last common ancestor of all life on Earth, proteins have been produced within cells via the decoding of codons written solely in the four-nucleotide genetic alphabet. We have now demonstrated the decoding of two new codons, written with an expanded genetic alphabet, and shown that they can be used to direct the site-specific incorporation of ncAAs into proteins. Remarkably, this reveals

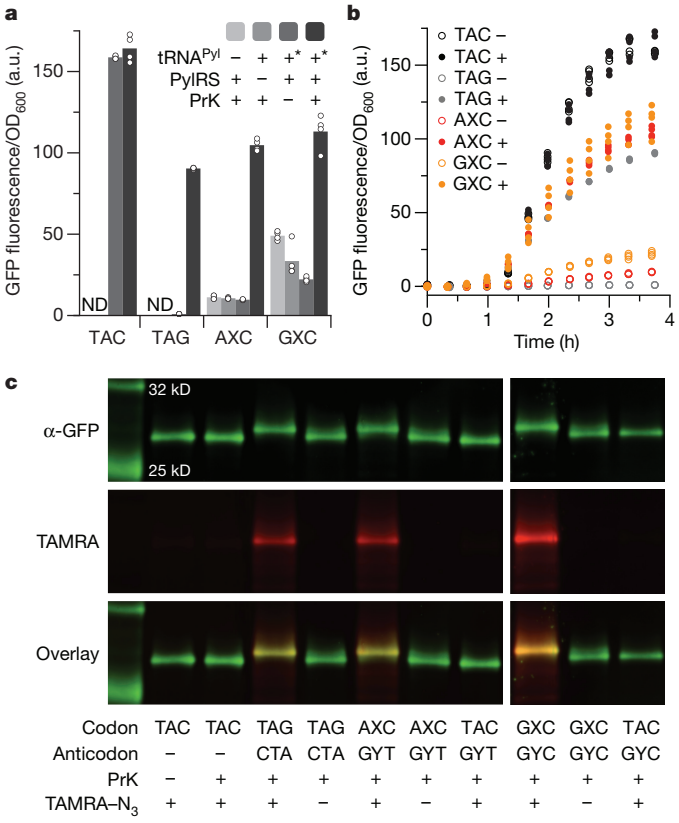


Figure 2 | Incorporation of PrK using PylRS–tRNA^{Pyl}. **a**, Fluorescence of cells expressing sfGFP with the indicated position 151 codon, in the presence (+) or absence (−) of a tRNA^{Pyl} with a cognate anticodon, PylRS, or 20 mM PrK in the medium, determined at the last time point in **b**. Asterisk denotes the absence of tRNA^{Pyl} in cells expressing sfGFP(TAC)^{151}; ND, not determined. Data shown as mean with individual values, $n = 4$ cultures, each propagated from an individual colony (biological replicates). **b**, Fluorescence timecourse of a subset of conditions in **a** (presence (+) or absence (−) of PrK in the medium). $t = 0$ corresponds to the addition of IPTG to induce expression of PylRS, T7 RNAP, and tRNA^{Pyl}; aTc was added at $t = 1$ h to induce expression of sfGFP. Individual data points shown from $n = 4$ cultures, each propagated from an individual colony (biological replicates). **c**, Western blots and fluorescence scans of sfGFP purified from cells expressing sfGFP and tRNA^{Pyl} (if present) with the indicated position 151 codon and anticodon, respectively, with or without conjugation of TAMRA or addition of PrK to the medium. sfGFP was purified from cultures collected at the last time point shown in **b**. For blot source data, see Supplementary Fig. 1. **d**, Relative abundance of amino acids at position 151 of sfGFP purified from cells expressing sfGFP(TAC)^{151} or sfGFP and tRNA^{Pyl} with the indicated position 151 codon and a cognate anticodon, respectively, as determined by LC–MS/MS and precursor ion intensity-based quantification; amino acids detected at <0.1% (on average, for all codons) are not shown. Data shown as mean with individual values, $n = 4$ purified sfGFP samples, each from a culture propagated from an individual colony, grown in the presence of PrK and collected at the last time point shown in **b**.

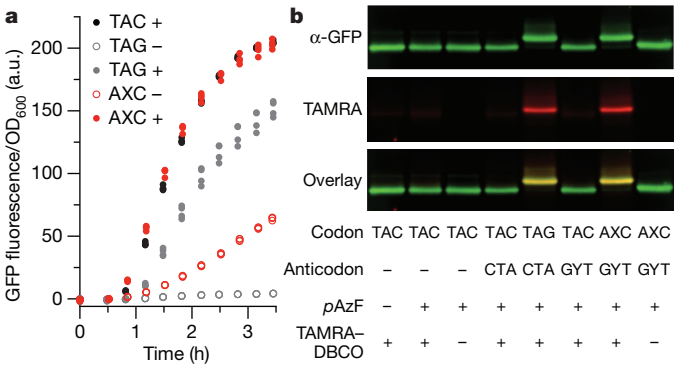


Figure 3 | Incorporation of *p*AzF using *p*AzFRS–tRNA^{pAzF}. **a**, Fluorescence of cells expressing sfGFP(TAC)^{151} or sfGFP and tRNA^{pAzF} with the indicated position 151 codon and a cognate anticodon, respectively, in the presence (+) or absence (−) of 5 mM *p*AzF in the medium. $t = 0$ corresponds to the addition of IPTG to induce expression of *p*AzFRS, T7 RNAP, and tRNA^{pAzF}; aTc was added at $t = 0.5$ h to induce expression of sfGFP. Individual data points shown from $n = 4$ cultures, each propagated from an individual colony (biological replicates). The fluorescence observed with sfGFP(AXC)^{151} in the absence of *p*AzF is attributed to charging of tRNA^{pAzF}(GYT) with a natural amino acid[16]. **b**, Western blot and fluorescence scan of sfGFP purified from cells expressing sfGFP and tRNA^{pAzF} (if present) with the indicated position 151 codon and anticodon, respectively, with or without conjugation of TAMRA or addition of *p*AzF to the medium. sfGFP was purified from cultures collected at the last time point shown in **a**. For blot source data, see Supplementary Fig. 1.

that for every step of information storage and retrieval, hydrogen bonds, so obviously central to the natural base pairs, may at least in part be replaced with complementary packing and hydrophobic forces. Despite their novel mechanism of decoding, the unnatural codons can be decoded as efficiently as their fully natural counterparts. We have examined the decoding of only two unnatural codons, but the UBP is unlikely to be limited to these, especially when combined with a recently reported Cas9 editing system that increases the range of sequences in which the UBP may be retained[2]; this would enable the exploration of more codons and further improve the fidelity of ncAA incorporation. Thus, the reported SSO is likely to be just the first of a new form of semi-synthetic life that is able to access a broad range of forms and functions not available to natural organisms.

Online Content Methods, along with any additional Extended Data display items and Source Data, are available in the online version of the paper; references unique to these sections appear only in the online paper.

Received 13 July; accepted 20 October 2017.

1. Leduc, S. *The Mechanisms of Life* (Rebman, 1911).
2. Zhang, Y. *et al.* A semisynthetic organism engineered for the stable expansion of the genetic alphabet. *Proc. Natl Acad. Sci. USA* **114,** 1317–1322 (2017).
3. Pédelacq, J. D., Cabantous, S., Tran, T., Terwilliger, T. C. & Waldo, G. S. Engineering and characterization of a superfolder green fluorescent protein. *Nat. Biotechnol.* **24,** 79–88 (2006).
4. Mukai, T., Lajoie, M. J., Englert, M. & Söll, D. Rewriting the genetic code. *Annu. Rev. Microbiol.* **71,** 557–577 (2017).
5. Liu, C. C. & Schultz, P. G. Adding new chemistries to the genetic code. *Annu. Rev. Biochem.* **79,** 413–444 (2010).
6. Shimizu, M., Asahara, H., Tamura, K., Hasegawa, T. & Himeno, H. The role of anticodon bases and the discriminator nucleotide in the recognition of some *E. coli* tRNAs by their aminoacyl-tRNA synthetases. *J. Mol. Evol.* **35,** 436–443 (1992).
7. Gorochowski, T. E., Avcilar-Kucukgoze, I., Bovenberg, R. A., Roubos, J. A. & Ignatova, Z. A minimal model of ribosome allocation dynamics captures trade-offs in expression between endogenous and synthetic genes. *ACS Synth. Biol.* **5,** 710–720 (2016).
8. Chatterjee, A., Sun, S. B., Furman, J. L., Xiao, H. & Schultz, P. G. A versatile platform for single- and multiple-unnatural amino acid mutagenesis in *Escherichia coli*. *Biochemistry* **52,** 1828–1837 (2013).

9. Nguyen, D. P. *et al.* Genetic encoding and labeling of aliphatic azides and alkynes in recombinant proteins via a pyrrolysyl-tRNA synthetase/tRNA(CUA) pair and click chemistry. *J. Am. Chem. Soc.* **131,** 8720–8721 (2009).
10. Voss, S. & Skerra, A. Mutagenesis of a flexible loop in streptavidin leads to higher affinity for the *Strep*-tag II peptide and improved performance in recombinant protein purification. *Protein Eng.* **10,** 975–982 (1997).
11. Hong, V., Presolski, S. I., Ma, C. & Finn, M. G. Analysis and optimization of copper-catalyzed azide–alkyne cycloaddition for bioconjugation. *Angew. Chem. Int. Ed.* **48,** 9879–9883 (2009).
12. Wandrey, G. *et al.* Probing unnatural amino acid integration into enhanced green fluorescent protein by genetic code expansion with a high-throughput screening platform. *J. Biol. Eng.* **10,** 11 (2016).
13. O'Donoghue, P., Ling, J., Wang, Y. S. & Söll, D. Upgrading protein synthesis for synthetic biology. *Nat. Chem. Biol.* **9,** 594–598 (2013).
14. Chin, J. W. *et al.* Addition of *p*-azido-L-phenylalanine to the genetic code of *Escherichia coli*. *J. Am. Chem. Soc.* **124,** 9026–9027 (2002).
15. Jewett, J. C., Sletten, E. M. & Bertozzi, C. R. Rapid Cu-free click chemistry with readily synthesized biarylazacyclooctynones. *J. Am. Chem. Soc.* **132,** 3688–3690 (2010).
16. Young, T. S., Ahmad, I., Yin, J. A. & Schultz, P. G. An enhanced system for unnatural amino acid mutagenesis in *E. coli*. *J. Mol. Biol.* **395,** 361–374 (2010).
17. Wang, K. *et al.* Optimized orthogonal translation of unnatural amino acids enables spontaneous protein double-labelling and FRET. *Nat. Chem.* **6,** 393–403 (2014).

Supplementary Information is available in the online version of the paper.

Acknowledgements We thank P. G. Schultz for the pEVOL-*p*AzF plasmid. This work was supported by the National Institutes of Health (GM118178 to F.E.R.). A.W.F. was supported by a National Science Foundation Graduate Research Fellowship (NSF/DGE-1346837).

Author Contributions Y.Z., J.L.P. and F.E.R. designed experiments. Y.Z. and J.L.P. designed plasmids. Y.Z., E.C.F. and J.L.P. performed and interpreted translation experiments. H.R.A. conducted and interpreted MS experiments. C.E.C. developed methods. A.W.F. and K.S.J. provided technical assistance. C.R.T. and F.E.R. provided project leadership. F.E.R. provided overall project supervision. Y.Z. and F.E.R. wrote the manuscript, with input from the other authors.

Author Information Reprints and permissions information is available at www.nature.com/reprints. The authors declare competing financial interests: details are available in the online version of the paper. Readers are welcome to comment on the online version of the paper. Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations. Correspondence and requests for materials should be addressed to F.E.R. (floyd@scripps.edu).

Reviewer Information *Nature* thanks A. Chatterjee, A. Deiters and P. Herdewyn for their contribution to the peer review of this work.

METHODS

No statistical methods were used to predetermine sample size. The experiments were not randomized and the investigators were not blinded to allocation during experiments and outcome assessment.

Materials. A complete list of plasmids and the sequences of oligonucleotides used in this work can be found in Supplementary Tables 1, 2. gBlock gene fragments and natural oligonucleotides (with standard purification and desalting) were purchased from IDT. Sequencing was performed by Genewiz. Plasmids were isolated using commercial miniprep kits (QIAprep, Qiagen or ZR Plasmid Miniprep Classic, Zymo Research). pEVOL-*p*AzF[14] was provided by P. Schultz.

Nucleosides of dNaM, dTPT3, NaM, TPT3, d5SICS and dMMO2^{bio} were synthesized (WuXi AppTec) and triphosphorylated (TriLink BioTechnologies LLC and MyChem LLC) commercially. All unnatural oligonucleotides were synthesized by Biosearch Technologies with purification by reverse phase cartridge.

Growth conditions. Unless otherwise stated, liquid bacterial cultures were grown in 300 μl of 2 × YT (casein peptone 16 g l^{-1}, yeast extract 10 g l^{-1}, NaCl 5 g l^{-1}) supplemented with potassium phosphate (50 mM, pH 7), referred to hereafter as medium, and incubated at 37 °C in 48-well flat-bottomed plates (CELLSTAR, Greiner Bio-One) with shaking at 200 r.p.m. Solid growth medium was prepared with 2% agar. Antibiotics were used, as appropriate, at the following concentrations: ampicillin, 100 μg ml^{-1}; zeocin, 50 μg ml^{-1}; chloramphenicol, 5 μg ml^{-1}. All selective agents were purchased commercially. Stock solutions of N^6-[(2-propynyloxy) carbonyl]-L-lysine HCl (PrK; SC-36420, Synchem) and *p*-azido-L-phenylalanine HCl (*p*AzF; 06162, Chem-Impex International) were dissolved in ddH_2O (1 M PrK) or 0.2 M NaOH (250 mM *p*AzF), respectively. Cell growth was measured using a Perkin Elmer Envision 2103 Multilabel Reader with a 590/20 nm filter. sfGFP fluorescence was measured using a Perkin Elmer Envision 2103 Multilabel Reader with a 485/14-nm filter for excitation and 535/25-nm filter for emission. The fluorescence of the medium used in each experiment was measured and subtracted from the fluorescence measurements of each cell culture.

Construction of synthetase expression plasmids. pGEX-*Mb*PylRS TetR was constructed by gene synthesis of a fragment corresponding to the sequence of the *M. barkeri* pyrrolysyl-tRNA synthetase (PylRS) and Gibson Assembly (Gibson Assembly Master Mix, New England Biolabs (NEB)) with a pGEX-4T-1 plasmid (Genscript), which removed the glutathione *S*-transferase gene and multiple cloning site from pGEX-4T-1 and placed *PylRS* under the control of $P_{tac\text{-}lacO}$. The resulting plasmid was then further modified by PCR and restriction digest cloning to remove the catabolite activator protein (CAP) binding site–P_{lac}–lacZα region and to introduce a fragment containing *tetR* expressed from a P_{bla} promoter (derived from pTETBHR2[18]) to yield pGEX-*Mb*PylRS TetR. pGEX-*Mjp*AzFRS TetR was cloned by introducing a T1 transcriptional terminator after *lacI* via PCR of pGEX-*Mb*PylRS TetR with phosphorylated primers (T4 PNK, NEB) with overhangs containing the T1 terminator sequence and blunt-ended ligation, replacement of P_{tac} with P_{lacUV5} by the same cloning strategy, and then PCR linearization of the resulting plasmid to exclude *PylRS* and introduce BglII and SalI restriction sites, which were then ligated with the BglII–SalI fragment containing *pAzFRS* from pEVOL-*p*AzF. pGEX-empty TetR, a plasmid used in Ser decoding experiments (to provide TetR) and used as a control plasmid in PrK incorporation experiments, was cloned by PCR linearization of pGEX-*Mb*PylRS TetR to exclude *PylRS*, followed by blunt-ended ligation.

Construction of sfGFP and tRNA expression plasmids. sfGFP and tRNA expression plasmids are based on the design of pAIO2X[2]. The *M. jannaschii* tRNA^{pAzF} and *M. mazei* tRNA^{Pyl} genes, as well as a small portion of the flanking 5′ leader and 3′ trailer sequences from *proK*, are derived from pULTRA[8]; the *E. coli serT* gene is derived from the native coding sequence. All sfGFP and tRNA expression plasmids were modified to replace the lacO controlling sfGFP expression with tetO, using phosphorylated primers with overhangs containing the tetO sequence and blunt-ended ligation.

To create the Golden Gate destination plasmids p[sfGFP(gg)^{151} tRNA^{Ser}(gg)], p[sfGFP(gg)^{151} tRNA^{Pyl}(gg)], and p[sfGFP(gg)^{151} tRNA^{pAzF}(gg)], which are used to introduce natural or unnatural position 151 sfGFP codons and tRNA anticodons, the lambda T_0 terminator of pAIO2X was removed via PCR and ligation, followed by a three-way Gibson Assembly of the resulting plasmid to simultaneously introduce a tRNA sequence (with a Golden Gate destination site) under the control of a T7 promoter and append a tandem C-terminal *Strep*-tag II[10] on *sfGFP* (also containing a Golden Gate destination site).

Expression plasmids p[sfGFP(TAG)^{151} tRNA^{Pyl}(CTA)], p[sfGFP(TAG)^{151} tRNA^{pAzF}(CTA)], and p[sfGFP(TAC)^{151} tRNA^{pAzF}(CTA)] were cloned by Golden Gate assembly (using natural inserts; see section Golden Gate assembly of UBP-containing plasmids). p[sfGFP(gg)^{151}], p[sfGFP(TAC)^{151}], and p[sfGFP(TAG)^{151}] were cloned by PCR linearization and ligation of a corresponding sfGFP and tRNA expression plasmid to remove the tRNA expression cassette. Owing to the presence of a BsaI restriction site near the *serT* anticodon when the anticodon is the amber suppressor CTA, p[sfGFP(TAG)^{151} tRNA^{Ser}(CTA)] was cloned by PCR linearization and ligation of p[sfGFP(TAG)^{151}] using primer overhangs to introduce the tRNA^{Ser}(CTA) expression cassette.

p[sfGFP(TAC)^{151} tRNA^{Pyl}(gg)] and p[sfGFP(TAC)^{151} tRNA^{pAzF}(gg)] were cloned by partial Golden Gate assembly (without the KpnI endonuclease and T5 exonuclease digest[2]) between the corresponding Golden Gate destination plasmid and a natural sfGFP insert (made using primer GFP151(TAC)), transformed into BL21(DE3), and selected on plates also containing 100 μM IPTG (to induce expression of sfGFP and thus verify the correct cloning of the sfGFP insert). Plasmids containing natural codon mutations at position 151 (that is, p[sfGFP(AGT)^{151}], p[sfGFP(ANC)^{151}] and p[sfGFP(GNC)^{151}]) were cloned by PCR amplification of p[sfGFP(TAC)^{151}] with primers containing these mutations and blunt-ended ligation. Similarly, plasmids containing natural anticodon mutations in *serT* (that is, p[sfGFP(gg)^{151} tRNA^{Ser}(GNT)]) were cloned by mutation of the CTA anticodon in p[sfGFP(TAG)^{151} tRNA^{Ser}(CTA)], followed by restriction digest cloning to replace sfGFP(TAG)^{151} with sfGFP(gg)^{151}.

Golden Gate assembly of UBP-containing plasmids. Plasmids containing UBPs in *sfGFP* (and if present, the gene encoding tRNA^{Ser}, tRNA^{Pyl}, or tRNA^{pAzF}) were generated by PCR amplification of template oligonucleotides containing dNaM and Golden Gate assembly[19], as previously described[2]; see Supplementary Table 2 for a full list of primers and templates and Supplementary Table 1 for their corresponding Golden Gate destination plasmids (annotations of plasmid sequences are provided in Supplementary Notes).

***In vivo* translation experiments.** Chemically competent YZ3 cells were prepared using $MgCl_2$ and $CaCl_2$, as previously described[20], but without the addition of glycerol, and transformed with synthetase expression plasmids (pGEX-*Mb*PylRS TetR or pGEX-*Mjp*AzFRS TetR) or an empty plasmid that expresses TetR (pGEX TetR, for the Ser incorporation experiments). Following transformation, single colonies were isolated and used to inoculate cultures that were then grown and then stored in glycerol (25% v/v) at −80 °C.

Electrocompetent YZ3 cells carrying synthetase expression plasmids or pGEX-empty TetR were prepared as previously described[2]. Freshly prepared electrocompetent YZ3 cells (50 μl) were electroporated with ~0.4 ng of Golden Gate assembled plasmids containing UBP(s) and then immediately diluted with 950 μl pre-warmed medium supplemented with chloramphenicol. A 5-μl aliquot of this dilution was then immediately diluted fivefold with the same pre-warmed medium, but additionally supplemented with dNaMTP (150 μM) and dTPT3TP (10 μM). Samples were incubated for 1 h at 37 °C without shaking and then various dilutions of the sample were plated on solid medium (2 ml in a 35 (D) × 10 (H) mm cell culture dish for each plate) supplemented with chloramphenicol, ampicillin, zeocin, dNaMTP (150 μM) and dTPT3TP (10 μM).

Following overnight growth (~12 h, 37 °C), individual colonies were used to inoculate liquid medium of the same composition as the solid medium. Cells were then monitored for growth, collected at $OD_{600} \approx 1$ and rapidly chilled to 4 °C (to pause growth, as individual cultures took varying amounts of time to reach $OD_{600} \approx 1$). Once all cultures were collected, cells were diluted to $OD_{600} \approx 0.1$–0.2 in 300 μl of medium and monitored for growth; any remaining culture not used in the dilution was pelleted and stored at −80 °C for subsequent plasmid isolation to determine UBP retention (*vide infra*). When cultures reached $OD_{600} \approx 0.4$–0.6, they were supplemented with NaMTP (250 μM) and TPT3TP (30 μM) (unless otherwise indicated; for example, Extended Data Fig. 4) and the appropriate ncAA (or ddH_2O for cultures without PrK or 0.2 M NaOH for cultures without *p*AzF) at the concentrations indicated in the respective figure captions. Cultures were then grown for 20 min before addition of 1 mM IPTG to induce T7 RNAP and transcription of tRNA^{Ser}, tRNA^{Pyl}, or tRNA^{pAzF}, and PylRS or *p*AzFRS, if present, and subsequently monitored for growth and fluorescence. Following a period of T7 RNAP and tRNA induction (0.5 h for Ser incorporation and *p*AzFRS experiments, 1 h for PylRS experiments), sfGFP was induced with 100 ng ml^{-1} anhydrotetracycline. After induction, cell cultures were rapidly cooled by shaking in a shallow ice water bath and collected at the time points indicated (see Figures). Cultures were aliquoted (50 μl for plasmid isolation to determine UBP retention, 50 μl for crude cell lysates, and 150 μl (Ser incorporation samples), 150 or 230 μl (PrK incorporation samples from cells expressing fully natural or unnatural codon-containing sfGFP, respectively), or 100 μl (*p*AzF incorporation samples) for affinity purification of sfGFP), pelleted and stored at −80 °C before being analysed as described below. Owing to the higher volume of culture (230 μl) used in the purification of some sfGFP samples in the PrK incorporation experiments, the crude cell lysates (for western blotting) of the corresponding samples were isolated from a separate experimental replicate.

Transformations with natural sfGFP plasmids were performed using freshly prepared chemically competent YZ3 cells carrying synthetase expression plasmids or pGEX-empty TetR (*vide supra*) and 1–10 ng natural sfGFP plasmid minipreps, with plating on solid medium. Following transformation, individual colonies were

inoculated, cultured, and processed as described above, with the exception that strains transformed with natural sfGFP plasmids in PrK (for example, Fig. 2) and *p*AzF (for example, Fig. 3) incorporation experiments were generally cultured without unnatural triphosphates (unless otherwise indicated, for example, Extended Data Fig. 4b). YZ3 cells transformed with natural sfGFP plasmids in Ser incorporation experiments (Fig. 1) were grown with the unnatural triphosphates (at the concentrations indicated above) in liquid medium only.

Statistics and reproducibility. Data presented in figures are representative datasets collected by single experimental replicates (unless otherwise indicated below) in which the colonies for each experimental condition were isolated from one transformation and plating, and were grown and processed in parallel; performing multiple transformations and platings and propagating several colonies from each transformation was cost-prohibitive owing to the amount of unnatural triphosphates required for culturing on solid medium. Additional experimental replicates performed by direct inoculation of transformations into liquid medium without plating, but under otherwise identical conditions, produced virtually identical results, as determined by fluorescence and growth (but without western blot or MS analysis, unless otherwise indicated).

Experimental replicates were performed as follows: Ser incorporation experiments (Fig. 1) and decoding experiments with near-cognate natural $tRNA^{Ser}$ (Extended Data Fig. 2) were run in parallel from the same transformation and were performed once with plating and once without. PrK incorporation experiments (Fig. 2, Extended Data Fig. 3, Extended Data Fig. 4c) and PrK titration experiments (Extended Data Fig. 5) were run in parallel from the same transformation (with the exception that the cell lysates analysed in the fourth, seventh, eighth and eleventh lanes of Extended Data Fig. 3a and the purified sfGFP samples analysed in the eighth and eleventh lanes of Fig. 2c were produced in a separate experiment) and were performed once with plating and once without. *p*AzF incorporation experiments (Fig. 3 and Extended Data Fig. 6) were performed once with plating and twice without. The unnatural ribotriphosphate control experiments (Extended Data Fig. 4a) were performed as separate experimental replicates from the main PrK incorporation experiments, were performed once with plating and once without, and were analysed by fluorescence, growth and western blot for both of these replicates. The experiments that correspond to Extended Data Figs 1 and 4b were performed once.

Western blots were performed on one cell lysate sample or one purified sfGFP sample from one experimental replicate. Technical replicates were performed as follows: Fig. 1e (one replicate), Fig. 2c (three replicates), Fig. 3b (two replicates), Extended Data Fig. 3a (one replicate), Extended Data Fig. 4a (one replicate), Extended Data Fig. 5a (two replicates).

In addition to the experimental replicates described above, experiments were also performed without plating and with minor technical variations (for example, growth in 96-well plates instead of 48-well plates, different induction schemes, and so on) and produced results consistent with the conclusions presented herein (assessed by fluorescence, growth, and western blot).

Cultures propagated from colonies transformed with different plasmids are necessarily separate transformations, whereas cultures grown under different medium conditions (for example, the presence or absence of an added amino acid or unnatural triphosphate) are derived from the same starter culture, that is, after the initial propagation of each colony to $OD_{600} \approx 1$ (*vide supra*), the dilution to $OD_{600} \approx 0.1$–0.2 in 300 μl was scaled according to the number of conditions to be tested, and the dilutions were performed as one batch and aliquoted.

Affinity purification of sfGFP. Cell pellets collected at the end of *in vivo* translation experiments were lysed by resuspension in 100 μl BugBuster (EMD Millipore) and incubation with shaking for 15 min at room temperature. Cell lysates were then diluted in Buffer W (50 mM HEPES pH 8, 150 mM NaCl, 1 mM EDTA) to a final volume equal to 500 μl minus the volume of beads, *vide infra*, and mixed with magnetic *Strep*-Tactin beads (5% (v/v) suspension of MagStrep 'type3' XT beads, IBA Lifesciences), equilibrated in Buffer W. One hundred microlitres of beads was used in the purification of all samples for which a yield is reported (Extended Data Table 3); all other samples were purified with 20 μl beads. Samples were then incubated for 30 min at 4 °C with constant gentle inversion. Beads were then pulled down with a magnetic rack, washed with Buffer W (2 × 500 μl), and eluted with 25 μl buffer BXT (100 mM Tris-HCl pH 8, 150 mM NaCl, 1 mM EDTA, 50 mM D-biotin; for PrK incorporation experiments, buffer BXT was made with 50 mM HEPES pH 8 instead of Tris, owing to the incompatibility of Tris with copper-catalysed click conjugation) for 10 min at room temperature with occasional vortexing. Purified proteins were quantified using a Qubit Protein Assay Kit (Thermo Fisher Scientific).

Sample preparation for LC–MS. Purified sfGFP proteins were dialysed (2,000 MWCO Slide-A-Lyze MINI dialysis cassettes, Thermo Fisher Scientific) with 2 × 20 min incubations in 2 l of 25 mM Tris-HCl pH 7.5 at room temperature, then quantified using a Qubit Protein Assay Kit. An aliquot corresponding to 3–3.5 μg protein was digested with a two-step protocol with lysyl endopeptidase (LysC; Wako Chemicals) and sequencing grade modified porcine trypsin (Promega) as described[21] with the following minor modifications: protein was dissolved in 15 μl solubilization buffer; heat denaturation was performed for 6 min at 95 °C; and the LysC digest was 2 h. Desalting of peptides was performed as described[22] with StageTips prepared in-house from 2 Empore disks C18 (3M Empore; Sigma-Aldrich) with a diameter of 1.60 mm. The resulting desalted peptides were lyophilized and then reconstituted in 12 μl of 2:3:7 by volume of 70% formic acid, 1-propanol, and 0.5% acetic acid. The concentration of the peptide stock solutions was estimated to be 100–125 ng μl^{-1} (assuming 50% recovery from the digestion and clean-up step). Peptide stock solutions were further diluted to a final peptide concentration of 2.85 ng μl^{-1} immediately before LC–MS analysis. MS samples were prepared in HPLC glass sample vials with a solution of 100 ng μl^{-1} sodium deoxycholate prepared in 0.1% TFA, 8% formic acid, and 5% DMSO as the diluent.

Liquid chromatography. Liquid chromatography was performed on an UltiMate 3500RS nanocapillary UHPLC (Thermo Fisher Scientific) equipped with a ProFlow flow module, a 10-port switching valve, a loading pump, and an autosampler operated at 7 °C. The trapping and analytical column were connected using a vented split setup[23] with a micro tee (IDEX) connected to the 10-port valve. The trapping column and analytical capillary column were prepared in-house using 360 μm OD and 150 μm ID fused silica capillaries (Molex) with a 2–3-mm-long Kasil frit prepared according to the instructions provided with the Frit Kit (Next Advance). The trapping and capillary column were slurry packed on a PC77 Pressure Injection Cell (Next Advance) to a length of 30 mm and 200 mm, respectively, using ReproSil-Pur 120 C18-AQ, 3 μm for the trapping column, and ReproSil-Pur 120 C18-AQ, 1.9 μm (Dr. Maisch GmbH) for the analytical column, respectively. Methanol was used as the packing solvent. All separations were performed at room temperature and the injection volume was 1 μl (2.85 ng peptides). Peptides were loaded onto the trap column at a flow rate of 5 μl min^{-1} with 2% ACN, 0.05% TFA as the solvent. Gradient elution of peptides was performed at a flow rate of 1 μl min^{-1} with the capillary pump. Eluent A consisted of 0.1% formic acid in water and eluent B was 0.1% formic acid in 80% acetonitrile, respectively. The following 90-min linear-gradient program was used: (min/%B) 0.0/2.0, 4.0/2.0, 5.0/8.0, 30.0/18.0, 45.0/24.0, 75.0/50.0, 78.0/60.0, 80.0/95.0, 83.0/95.0, 85.0/2.0, 90.0/2.0. A control injection consisting of 40 fmol of a tryptic digest of BSA was performed between samples for quality control purposes. Care was taken to eliminate carryover between sample injections. Sample injections were followed by four injections with 2 μl MS solvent followed by a solvent blank, which was used to assess carryover before the injection of the next sample. This procedure effectively eliminated carryover between samples originating from different experimental conditions.

Mass spectrometry. Nanocapillary LC–MS/MS was performed on an Impact II QqTOF mass spectrometer (Bruker) as described[24] with the following modifications. The instrument was equipped with a captive spray ion source fitted with a 20-μm ID spray tip and the nanoBooster dopant system for increased sensitivity. The ion source was operated with a spray voltage of 1,000 V at 150 °C with nitrogen (4 l min^{-1}) as the drying gas. The nanoBooster was operated at 0.2 bar with a solvent mixture of 3:1 by volume of acetonitrile and methanol. Calibration of the mass spectrometer was performed daily using a sodium-TFA solution prepared as described[25]. Spectra were collected at a resolving power of 60,000 using a data-dependent acquisition method with a cycle time of 1.1 s. Each scan event consisted of a single precursor ion scan (scan range 150 Da to 2,200 Da, 5 Hz) followed by 20 scheduled tandem MS scans performed at 20 Hz. Collision-activated dissociation was performed with a precursor isolation window of 2 Da and using nitrogen as the collision gas. The collision energy was dynamically adjusted between 23 and 65 eV depending on ion mass and charge state of the selected precursor as defined in the acquisition software (Bruker Compass 1.9 Otof Control Version 4.0, Build 60.11, Top20 Auto MS MS method). Dynamic exclusion parameters were as follows: the strict active exclusion option was activated, the preferred precursor ion charge range was 2–6, singly charged precursors were excluded, and acquired precursor ions were excluded for 0.4 min unless the precursor ion intensity exceeded 3× the intensity of the precursor observed in the previous MS1 scan. All data were collected in profile mode.

Bioinformatics. *Database searching.* Database searching and identification of peptides containing natural amino acids and ncAAs was performed according to a previously described workflow[26]. Database searches were performed with MaxQuant[27] v. 1.5.8.0 using the default search parameters with the following minor changes. The databases consisted of a custom database of twenty sfGFP variants representing all natural amino acids at position 151, and an *E. coli* reference proteome database with 4,306 protein entries (UniProt, Strain MG1655 retrieved on 16 August 2016). The minimum peptide length was 5; fixed modifications were carbamidomethyl (C); variable modifications were deamidation (N/Q), oxidation (M), and PrK, which represents the unnatural amino acid (N^6-[(2-propynyloxy)carbonyl]-L-lysine) and is defined as a custom modification ($+C_8H_8O$, +96.0575 Da)

to Asn. Peptide spectral matches and protein identifications were reported with a false discovery rate of 1% each. Proteins and peptides matching the contaminant database were removed by filtering in Microsoft Excel. A summary of all identified reporter peptides and three sfGFP reference peptides is presented in Extended Data Table 4.

Reporter peptide quantification in Skyline. LC–MS data are reported in Extended Data Table 4. Quantification of LC–MS chromatographic peak intensities was performed by extracting MS1 precursor ion chromatograms with Skyline Software (v. 3.6.0.10167)[28]. All reporter peptides identified during database searching were imported into Skyline along with three reference sfGFP peptides that were used for normalization of reporter peptide intensities. All peptides used for quantification are listed in Extended Data Table 4. Extracted precursor ion chromatograms were generated for the most abundant charge state for each peptide. Chromatographic peak integration was manually verified to ensure consistent peak integration across all samples. Further data processing was performed in Microsoft Excel. Reporter peptide intensities were normalized by dividing the reporter peptide peak areas for each sample by the sum of all three reference peptide peak areas from the same sample, which enabled direct comparison of reporter peptide intensities across samples. The normalized reporter peptide intensities corresponding to each amino acid detected at position 151 of sfGFP were summed and divided by the total normalized peak area for all observed reporter peptides within each sample to obtain the extracted precursor ion intensities presented in the main text, Figs 1f, 2d, and Extended Data Table 1. Finally, these relative percentages were summed for each amino acid across replicates and used to calculate a 95% confidence interval to assess the statistical significance of the results.

Click conjugation of TAMRA. For copper-catalysed click conjugation of TAMRA, 200 ng purified sfGFP protein (3–5 μl, depending on the concentration of the eluted protein following purification) was incubated with 2 mM tris(3-hydroxypropyltriazolylmethyl)amine (THPTA) (Sigma-Aldrich), 1 mM $CuSO_4$, 15 mM sodium ascorbate (Sigma-Aldrich) and 0.5 mM TAMRA-azide (Click Chemistry Tools; product #AZ109) for 1 h at room temperature in darkness. For copper-free click conjugation of TAMRA, 200 ng purified sfGFP protein (3–5 μl, depending on the concentration of the eluted protein following purification) was incubated with 0.1 mM TAMRA-DBCO (Click Chemistry Tools; product #A131) overnight at room temperature in darkness. Following click conjugation, all steps were performed with care to minimize the exposure of the samples, gels, and blots to ambient light to minimize photobleaching of TAMRA.

Western blotting of TAMRA-conjugated sfGFP. Two hundred nanograms of purified and conjugated sfGFP protein was mixed 2:1 (v:v) with loading buffer (250 mM Tris-HCl, 30% (v/v) glycerol, 2% (w/v) SDS), heated for 5 min at 95 °C and subjected to SDS–PAGE (stacking gel: 5% (w/v) acrylamide:bis-acrylamide 29:1 (Fisher), 0.125 M Tris-HCl and 0.1% SDS, pH 6.8 (ProtoGel Stacking Buffer, National Diagnostics); resolving gel: 15% (w/v) acrylamide:bis-acrylamide 29:1 (Fisher), 0.375 M Tris-HCl and 0.1% SDS, pH 8.8 (ProtoGel Resolving Buffer, National Diagnostics; 1.5 mm Mini-PROTEAN Short Plates (Bio-Rad)) with a protein ladder (Colour Prestained Protein Standard, Broad Range, NEB). Gels were run at 50 V for 15 min and then at 120 V for ~5 h in SDS–PAGE buffer (25 mM Tris base, 200 mM glycine, 0.1% (w/v) SDS).

Following SDS–PAGE, samples were transferred to low-fluorescence PVDF (0.2 μm, BioRad) by wet transfer (20% (v/v) MeOH, 50 mM Tris base, 400 mM glycine, 0.0373% (w/v) SDS; 90 V, 1 h, 4 °C). Membranes were blocked overnight at 4 °C in 5% (w/v) nonfat milk in PBS-T (PBS pH 7.4, 0.01% (v/v) Tween-20), probed with a rabbit anti-GFP antibody (product #G1544, lot 046M4871V, Sigma-Aldrich; 1:2,000 in PBS-T) for 1 h at room temperature, washed with PBS-T (1 × 5 min), probed with a goat anti-rabbit Alexa Fluor 647-conjugated antibody (product #A32733, lot #SD250298, Thermo Fisher Scientific; 1:20,000 in PBS-T) for 45 min at room temperature, washed with PBS-T (3 × 5 min), and subsequently imaged using a flatbed laser scanner (Typhoon 9410 with Typhoon Scanner Control v5 (Build 410 5.0.0409.0700), GE Healthcare Life Sciences); 50-μm resolution; 532-nm laser excitation and 580/30-nm emission filter with 400 V PMT for TAMRA; 633-nm laser excitation and 670/30-nm emission filter with 500 V PMT for Alexa Fluor 647). Images were pseudocoloured and quantified using Image Studio Lite (v. 5.2.5, LI-COR Biosciences). Overlays of the TAMRA and Alexa Fluor 647 scans were produced using Paint.NET (v. 3.5.10). Western blots in every multi-blot panel (that is, Fig. 2c, Extended Data Figs 3a, 4a, 5a) were performed, imaged, and processed in parallel under identical conditions.

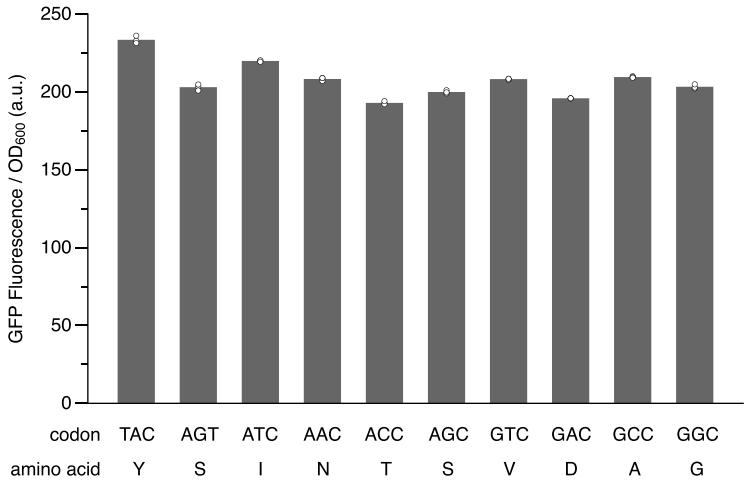
Western blotting of cell lysates. Cell pellets collected at the end of *in vivo* translation experiments were resuspended in ddH_2O to a calculated $OD_{600} \approx 2$ (based on the OD_{600} of cultures at the time of collection). Five microlitres of cells was then mixed with 4 μl loading buffer (250 mM Tris-HCl, 30% (v/v) glycerol, 2% (w/v) SDS, 6.25% (v/v) βME) and 1 μl loading dye (product #B7024S, NEB), heated for 5 min at 95 °C and subjected to SDS–PAGE under identical conditions as above, with the exception that gels were run at 50 V for 15 min and then at 120 V for ~2 h.

Following SDS–PAGE, samples were transferred to PVDF (0.45 μm Immobilon, EMD Millipore) and western blotting was performed as above, with the exception that the membrane was probed with a goat anti-rabbit HRP (Bio-Rad product #170-6515, lot #64106148) secondary antibody (instead of the goat anti-rabbit Alexa Fluor 647 antibody) for 1 h at room temperature, washed with PBS-T (3 × 5 min), incubated with chemiluminescent substrate (Pierce ECL Western Blotting Substrate, Thermo Fisher Scientific) for 5 min, and imaged with a ChemiDoc-It Imager (UVP) and UVP VisionWorks LS (v 8.6.15114.8618) software. Multiple exposures were taken and images used in the paper were processed in UVP VisionWorks, using the noise removal setting (Starfield Subtraction), and then further processed (colour inverted) in Image Studio Lite (v. 5.2.5, LI-COR Biosciences).

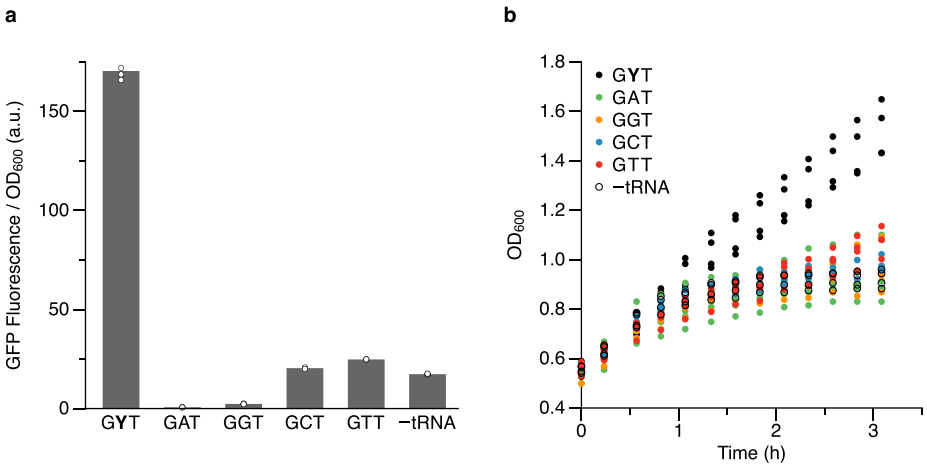
Biotin shift assay. The retention of the UBP(s) in isolated plasmids was determined as previously described[2] using d5SICSTP and $dMMO2^{bio}TP$, and the primers indicated in Supplementary Table 2. Images were quantified using Image Studio Lite (v. 5.2.5, LI-COR Biosciences). UBP retention was normalized by dividing the percentage raw shift of each plasmid sample by the percentage raw shift of the dNaM-containing oligonucleotide template used in the assembly of the input plasmid, except for UBP retentions in the gene encoding $tRNA^{pAzF}$, which are reported as raw shifts. This was due to an inability to correctly amplify the sequence containing the $tRNA^{pAzF}$ anticodon (under the conditions used for the PCR in the biotin shift assay) with primers that are compatible with both the control oligonucleotide and plasmids. Thus, primers that anneal outside the $tRNA^{pAzF}$ sequence (which are incompatible with the control oligonucleotide) were used in the PCR for the biotin shift assay. No such difficulty was encountered under the conditions used to generate the dNaM–dTPT3 insert used in plasmid assembly, and thus the problem appears to be specific to the PCR conditions used in the biotin shift assay.

Data availability. The data that support the findings of this study are available from the corresponding author upon reasonable request. Plasmid sequences have been deposited in the GenBank database under the accession numbers MF996900–MF996913.

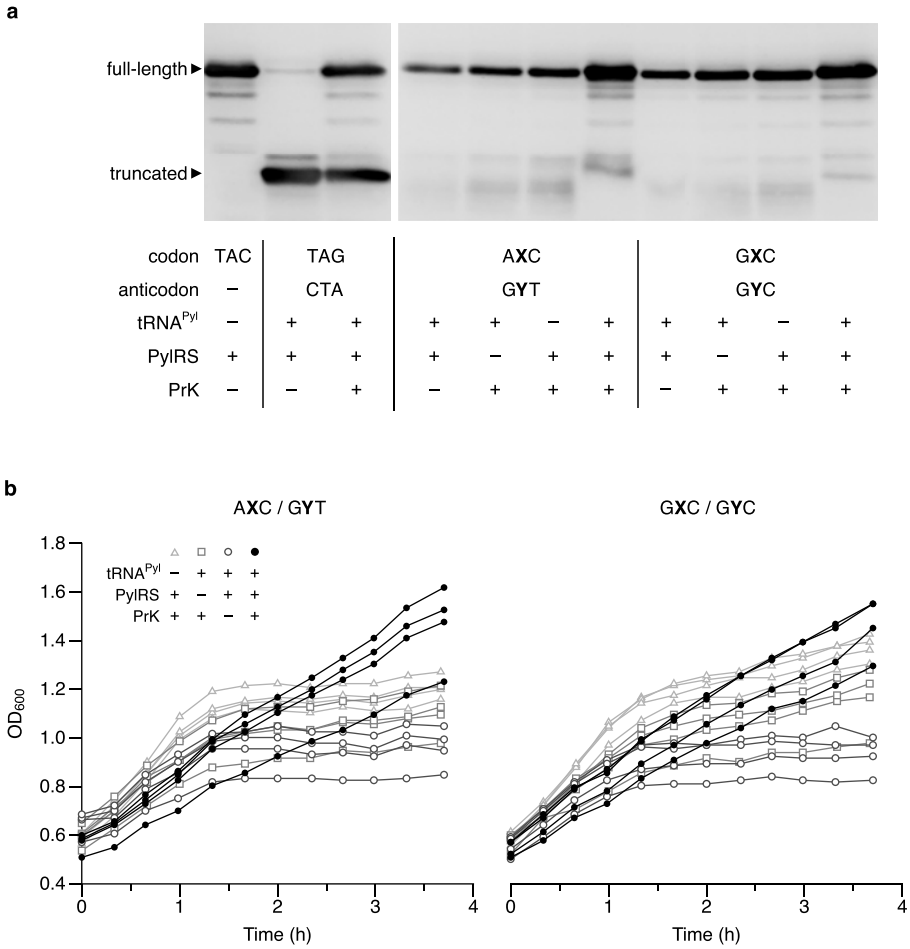
18. Smith, P. A. & Romesberg, F. E. Mechanism of action of the arylomycin antibiotics and effects of signal peptidase I inhibition. *Antimicrob. Agents Chemother.* **56,** 5054–5060 (2012).
19. Engler, C., Kandzia, R. & Marillonnet, S. A one pot, one step, precision cloning method with high throughput capability. *PLoS One* **3,** e3647 (2008).
20. Chan, W. T., Verma, C. S., Lane, D. P. & Gan, S. K. A comparison and optimization of methods and factors affecting the transformation of *Escherichia coli*. *Biosci. Rep.* **33,** e00086 (2013).
21. Pirman, N. L. *et al.* A flexible codon in genomically recoded *Escherichia coli* permits programmable protein phosphorylation. *Nat. Commun.* **6,** 8130 (2015).
22. Rappsilber, J., Mann, M. & Ishihama, Y. Protocol for micro-purification, enrichment, pre-fractionation and storage of peptides for proteomics using StageTips. *Nat. Protocols* **2,** 1896–1906 (2007).
23. Link, A. J., Jennings, J. L. & Washburn, M. P. Analysis of protein composition using multidimensional chromatography and mass spectrometry. *Curr. Prot. Protein Sci.* **78,** 23.21.21–23.21.25 (2001).
24. Beck, S. *et al.* The Impact II, a very high-resolution quadrupole time-of-flight instrument (QTOF) for deep shotgun proteomics. *Mol. Cell. Proteomics* **14,** 2014–2029 (2015).
25. Moini, M., Jones, B. L., Rogers, R. M. & Jiang, L. Sodium trifluoroacetate as a tune/calibration compound for positive- and negative-ion electrospray ionization mass spectrometry in the mass range of 100–4000 Da. *J. Am. Soc. Mass Spectrom.* **9,** 977–980 (1998).
26. Aerni, H. R., Shifman, M. A., Rogulina, S., O'Donoghue, P. & Rinehart, J. Revealing the amino acid composition of proteins within an expanded genetic code. *Nucleic Acids Res.* **43,** e8 (2015).
27. Cox, J. & Mann, M. MaxQuant enables high peptide identification rates, individualized p.p.b.-range mass accuracies and proteome-wide protein quantification. *Nat. Biotechnol.* **26,** 1367–1372 (2008).
28. MacLean, B. *et al.* Skyline: an open source document editor for creating and analyzing targeted proteomics experiments. *Bioinformatics* **26,** 966–968 (2010).



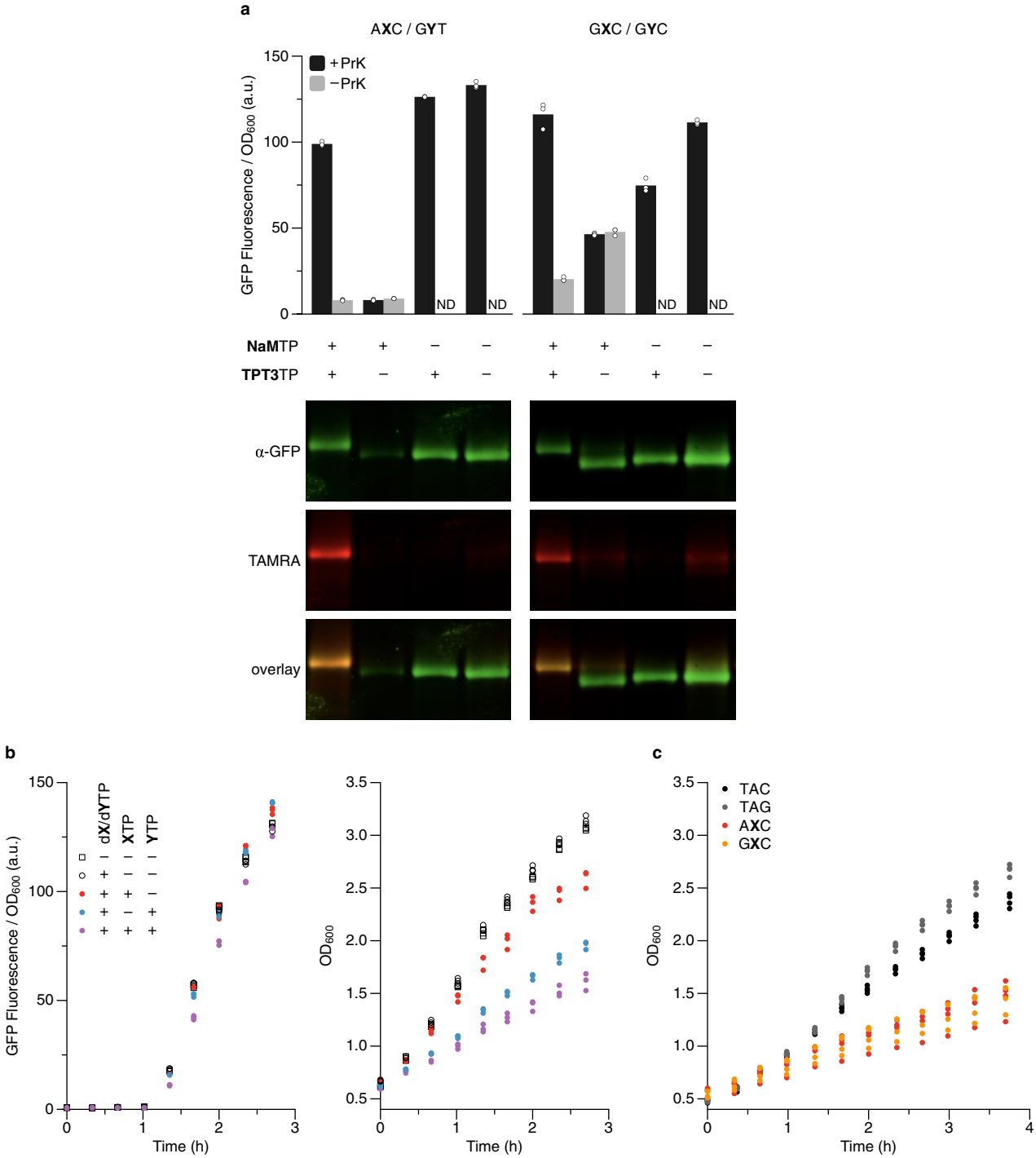
Extended Data Figure 1 | Fluorescence of cells expressing sfGFP with various codons at position 151. Strain YZ3 cells carrying a plasmid expressing TetR and an sfGFP plasmid with the indicated position 151 codon were grown to an OD_{600} of approximately 0.5 and induced with IPTG and aTc. Fluorescence measurements were taken after 3 h of induction. Data shown as mean with individual values, $n = 3$ cultures originating from the same colony and grown in parallel (technical replicates).



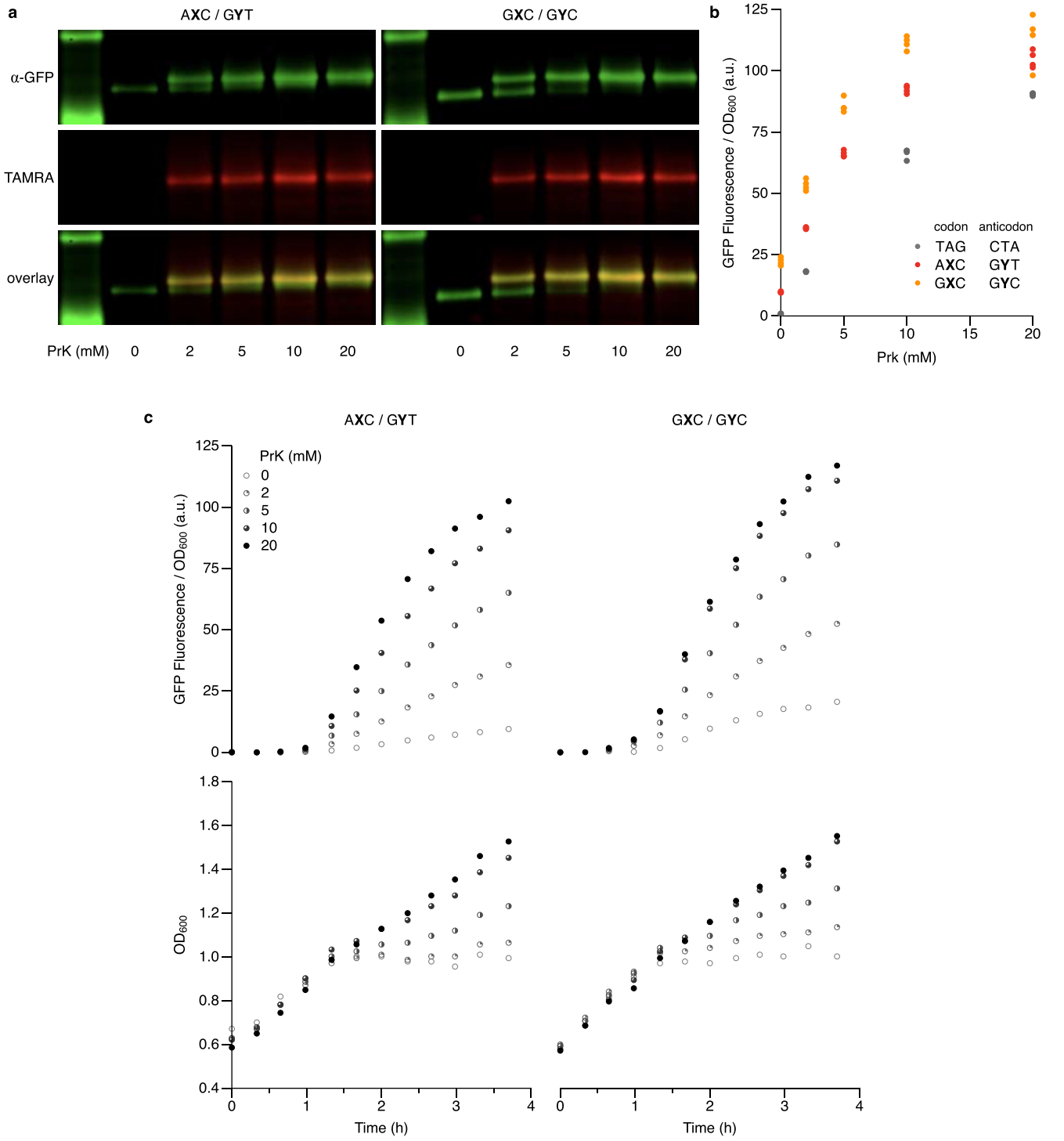
Extended Data Figure 2 | Decoding of the AXC codon with natural near-cognate anticodons. a, **b**, Fluorescence (**a**) and growth (**b**) of cells expressing sfGFP(AXC)[151] with or without $tRNA^{Ser}$ with the indicated anticodon. Cells were induced as described in Fig. 1c, d and fluorescence measurements correspond to the last time point shown in Fig. 1c. Values for the GYT anticodon and in the absence of $tRNA^{Ser}$ (–tRNA) correspond to the same values in Fig. 1c, d. Individual data points (with means in **a**) shown from $n = 4$ cultures, each propagated from an individual colony (biological replicates).



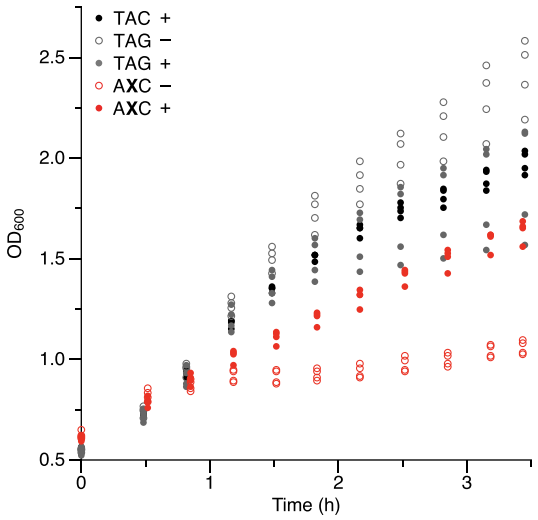
Extended Data Figure 3 | Western blots and growth of cells decoding AXC and GXC codons with tRNA^{Pyl}. a, Western blot of lysates (normalized by OD_{600}) from cells expressing sfGFP with the indicated position 151 codon, in the presence (+) or absence (–) of a tRNA^{Pyl} with a cognate anticodon, PylRS, or 20 mM PrK in the medium. Blots were probed with an anti-GFP antibody (N-terminal epitope). Cells were induced and collected at an equivalent time point, as described in Fig. 2b. For blot source data, see Supplementary Fig. 1. **b**, Growth of cultures analysed in Fig. 2a. The fold change in OD_{600} between induction of sfGFP ($t = 1$ h) and the final time point is greatest when all components necessary for aminoacylating tRNA^{Pyl} are present. Variations in the absolute value of OD_{600} are due to small variations in cell density at the start of T7 RNAP (and, if present, tRNA^{Pyl}) induction ($t = 0$). Individual data points shown from $n = 4$ cultures, each propagated from an individual colony (biological replicates).



Extended Data Figure 4 | Decoding of AXC and GXC codons with tRNA^{Pyl} and cell growth as a function of added unnatural ribotriphosphates. a, Fluorescence (upper panel) and western blots of purified sfGFP (lower panel) from cells expressing sfGFP and tRNA^{Pyl} with the position 151 codon/anticodon indicated, in the presence (+) or absence (–) of each unnatural ribotriphosphate in the medium, and with or without 20 mM PrK. Cells were induced as described in Fig. 2b and fluorescence measurements were taken at the end of induction (~3.5 h), before collecting the cells and purifying the sfGFP protein for click conjugation of TAMRA and western blotting. Western blots were probed with an anti-GFP antibody and imaged to detect both sfGFP and the conjugated TAMRA; all lanes correspond to sfGFP purified from cells grown with added PrK. Data shown as mean with individual values, $n = 3$ cultures, each propagated from an individual colony (biological replicates); ND, not determined. For blot source data, see Supplementary Fig. 1. **b**, Fluorescence and growth of cells expressing sfGFP(TAC)^{151} in the presence (+) or absence (–) of both unnatural deoxyribotriphosphates and each unnatural ribotriphosphate. $t = 0$ corresponds to the addition of IPTG to induce expression of T7 RNAP; aTc was added at $t = 1$ h to induce expression of sfGFP. Individual data points shown from $n = 3$ cultures, each propagated from an individual colony (biological replicates). At the concentrations used (see Methods), dNaMTP and dTPT3TP do not inhibit cell growth, whereas both unnatural ribotriphosphates, particularly TPT3TP, cause some inhibition of growth. **c**, Cell growth corresponding to the cultures with added PrK (20 mM) whose fluorescence is shown in Fig. 2b. Cells expressing sfGFP with natural codons were grown without any unnatural triphosphates, whereas cells expressing sfGFP with unnatural codons were grown with both unnatural deoxy- and ribotriphosphates. Individual data points shown from $n = 4$ cultures, each propagated from an individual colony (biological replicates).



Extended Data Figure 5 | Decoding of AXC and GXC codons with tRNA^{Pyl} as a function of PrK concentration in the medium. a, Western blots of sfGFP purified from cells expressing sfGFP and tRNA^{Pyl} with the indicated position 151 codon/anticodon, with conjugation of TAMRA and the addition of PrK to the medium at the indicated concentrations. sfGFP was induced and purified from cells collected as described in Fig. 2b. Western blots were probed with an anti-GFP antibody and imaged to detect both sfGFP and the conjugated TAMRA. For blot source data, see Supplementary Fig. 1. **b**, Fluorescence of cells (measured at the last time point shown in **c**) expressing sfGFP and tRNA^{Pyl} with the indicated position 151 codon and anticodon, respectively, as a function of PrK concentration in the medium. Fluorescence values for the 0 and 20 mM PrK conditions are the same as the (–) and (+) PrK conditions, respectively, shown in Fig. 2b. Individual data points shown from $n = 4$ cultures, each propagated from an individual colony (biological replicates). **c**, Timecourse analysis of fluorescence and cell growth in **b**. For clarity, only one representative culture (of four) is shown for each codon/anticodon pair and PrK concentration. We attribute the low level of sfGFP produced in the absence of PrK to decoding by endogenous tRNAs and loss of UBP retention in *sfGFP* (Extended Data Table 2). However, the relative amount of sfGFP that contains PrK (**a**) and absolute amount of sfGFP expressed (**b**, **c**) increased in a dose-dependent manner with increasing PrK in the medium, ultimately resulting in nearly full incorporation of PrK, suggesting that endogenous read-through of the AXC and GXC codons can be efficiently suppressed with sufficient concentrations of charged PrK-tRNA^{Pyl}(GYT) or PrK-tRNA^{Pyl}(GYC).



Extended Data Figure 6 | Cell growth of the cultures whose fluorescence is shown in Fig. 3a. Individual data points shown from $n = 4$ cultures, each propagated from an individual colony (biological replicates).

Extended Data Table 1 | Relative abundance of amino acids at position 151 in sfGFP for experiments described in Figs 1f and 2d

Sample	Relative precursor ion intensities (%)									
	S	Y	PrK	I/L	N	V	K	G	C	M
sfGFP(AGT)^{151}	99.80	0.03	0.06	0.00	0.04	0.03	0.00	0.02	0.02	0.00
sfGFP(A**X**C)^{151} / tRNA^{Ser}(G**Y**T)	98.47	0.04	0.04	1.23	0.14	0.02	0.00	0.05	0.01	0.00
sfGFP(TAC)^{151}	0.11	99.71	0.06	0.00	0.05	0.02	0.00	0.02	0.02	0.01
sfGFP(TAG)^{151} / tRNA^{Pyl}(CTA)	0.06	0.04	99.53	0.00	0.04	0.01	0.29	0.01	0.01	0.00
sfGFP(A**X**C)^{151} / tRNA^{Pyl}(G**Y**T)	0.25	0.03	96.16	2.06	1.06	0.02	0.37	0.03	0.01	0.00
sfGFP(G**X**C)^{151} / tRNA^{Pyl}(G**Y**C)	0.06	0.04	97.50	0.00	0.01	1.26	0.74	0.37	0.01	0.00

Sample	95% CI (%)									
	S	Y	PrK	I/L	N	V	K	G	C	M
sfGFP(AGT)^{151}	0.31	0.04	0.09	0.00	0.06	0.05	0.01	0.03	0.03	0.00
sfGFP(A**X**C)^{151} / tRNA^{Ser}(G**Y**T)	0.73	0.04	0.03	0.64	0.04	0.01	0.00	0.04	0.01	0.00
sfGFP(TAC)^{151}	0.06	0.11	0.05	0.00	0.03	0.02	0.00	0.01	0.02	0.00
sfGFP(TAG)^{151} / tRNA^{Pyl}(CTA)	0.03	0.02	0.11	0.00	0.02	0.02	0.03	0.01	0.01	0.00
sfGFP(A**X**C)^{151} / tRNA^{Pyl}(G**Y**T)	0.13	0.02	0.25	0.06	0.03	0.01	0.06	0.01	0.02	0.01
sfGFP(G**X**C)^{151} / tRNA^{Pyl}(G**Y**C)	0.05	0.04	0.70	0.00	0.01	0.24	0.28	0.22	0.01	0.00

sfGFP purified from cells expressing sfGFP with or without tRNAs with the indicated position 151 codon and anticodon, respectively, were analysed by LC–MS/MS. The extracted precursor ion intensities for the reporter peptides LEYNFNSHNVX^{151}ITADK (X denotes PrK or any identified natural amino acid except K or R) and LEYNFNSHNVX^{151} (if X is K or R) are expressed as a percentage of the sum of ion intensities for all observable reporter peptides. The table of values corresponds to the mean relative abundances and 95% confidence intervals of all amino acids detected at position 151 of sfGFP, $n = 4$ purified sfGFP samples, each from a culture propagated from an individual colony. Values <0.1% (on average, for the codons indicated in the respective main text figures) are excluded from the data presented in Figs 1f and 2d.

Extended Data Table 2 | UBP retention

aaRS	tRNA	**NaM**TP	**TPT3**TP	Codon	UBP retention in *sfGFP* (%)	Anti-codon	UBP retention in tRNA gene (%)
SerRS	–	+	+	A**X**C	98±0	–	n/a
SerRS	Ser	+	+	A**X**C	98±2	G**Y**T	89±2
SerRS	Ser	+	+	A**X**C	94±8	GAT	n/a
SerRS	Ser	+	+	A**X**C	94±2	GGT	n/a
SerRS	Ser	+	+	A**X**C	95±0	GCT	n/a
SerRS	Ser	+	+	A**X**C	95±1	GTT	n/a
–	Pyl	+	+	A**X**C	97±1	G**Y**T	89±2
PylRS	–	+	+	A**X**C	97±1	–	n/a
PylRS	Pyl	+	+	TAC	n/a	G**Y**T	92±3
PylRS	Pyl	+	+	A**X**C	96±1	G**Y**T	90±2
PylRS	Pyl	+	+	A**X**C	98±0	G**Y**T	95±2
PylRS	Pyl	+	–	A**X**C	98±1	G**Y**T	96±1
PylRS	Pyl	–	+	A**X**C	98±1	G**Y**T	95±1
PylRS	Pyl	–	–	A**X**C	97±1	G**Y**T	94±4
–	Pyl	+	+	G**X**C	98±1	G**Y**C	96±3
PylRS	–	+	+	G**X**C	97±3	–	n/a
PylRS	Pyl	+	+	TAC	n/a	G**Y**C	96±1
PylRS	Pyl	+	+	G**X**C	97±1	G**Y**C	95±0
PylRS	Pyl	+	+	G**X**C	96±3	G**Y**C	97±1
PylRS	Pyl	+	–	G**X**C	96±2	G**Y**C	97±1
PylRS	Pyl	–	+	G**X**C	97±2	G**Y**C	97±0
PylRS	Pyl	–	–	G**X**C	96±1	G**Y**C	97±1
*p*AzFRS	*p*AzF	+	+	A**X**C	98±0	G**Y**T	90±1
*p*AzFRS	*p*AzF	+	+	TAC	n/a	G**Y**T	91±1

Retention of the UBP(s) in plasmids with the indicated position 151 codons of sfGFP and anticodons of the indicated tRNAs were determined for a time point before sfGFP induction and at the end of induction, as described in Methods. The reported values are the mean UBP retention over the course of the induction (calculated from the retentions at these two time points) ± 95% CI, $n = 4$ cultures, each propagated from an individual colony, except for values in rows shaded grey, for which $n = 3$. n/a, not applicable (because the relevant sequence is natural or absent). All plasmids were isolated from cultures grown in the presence of 20 mM PrK or 5 mM *p*AzF (except for Ser incorporation experiments). SerRS indicates charging with the endogenous *E. coli* synthetase. Minus sign denotes the absence of PylRS in cells with $tRNA^{Pyl}$ or the absence of an ectopically expressed tRNA. Retentions in rows shaded green correspond to cultures from which sfGFP was also purified and analysed by LC–MS/MS and/or western blot of TAMRA-conjugated sfGFP (see Figs 1f (Ser), 2d (PrK), and 3b (*p*AzF)); rows shaded grey correspond to the cultures analysed in Extended Data Fig. 4. Despite the fact that all four unnatural triphosphates enter the cell through the same transporter and thus competitively inhibit one another's entry, no differences in UBP retention were observed with the presence (+) or absence (–) of NaMTP and/or TPT3TP in the medium. These data, and the requirement for both unnatural ribotriphosphates for high levels of sfGFP expression with high-fidelity PrK incorporation (Extended Data Fig. 4), collectively demonstrate that the expression level of *Pt*NTT2 in strain YZ3[2] imports the requisite levels of unnatural triphosphates necessary to sustain UBP replication and transcription.

Extended Data Table 3 | Yields of sfGFP protein expressed in Ser, PrK and *p*AzF incorporation experiments

Sample	aaRS	Yield (μg ml^{-1})	Relative to control (%)	Total fluor (a.u.)	Relative to control (%)
sfGFP(AGT)^{151}	SerRS (endogenous)	100±8	100	269	100
sfGFP(A**X**C)^{151} / tRNA^{Ser}(G**Y**T)		97±9	96	259	96
sfGFP(TAC)^{151}	PylRS	135±17	100	400	100
sfGFP(TAG)^{151} / tRNA^{Pyl}(CTA)		87±6	65	242	60
sfGFP(A**X**C)^{151} / tRNA^{Pyl}(G**Y**T)		54±4	40	153	38
sfGFP(G**X**C)^{151} / tRNA^{Pyl}(G**Y**C)		55±6	41	166	41
sfGFP(TAC)^{151}	*p*AzFRS	127±15	100	405	100
sfGFP(TAG)^{151} / tRNA^{pAzF}(CTA)		75±9	59	287	71
sfGFP(A**X**C)^{151} / tRNA^{pAzF}(G**Y**T)		86±6	68	333	82

Yields were calculated from the total amount of protein purified and the volume of culture used for purification (see Methods). Data are mean ± s.d. ($n = 4$ sfGFP samples, each purified from a culture propagated from an individual colony) and were determined from the same cultures analysed in Figs 1f (for SerRS) and 2d (for PylRS), as well as the cultures corresponding to the (+) *p*AzF samples in Fig. 3a (for *p*AzFRS). Yields of purified sfGFP are comparable to the mean total fluorescence (not normalized to OD_{600}) of the cultures from which they were purified. Fluorescence values correspond to the time point at which cells were collected for sfGFP purification; see Figs 1c (Ser), 2b (PrK), and 3a (*p*AzF).

Extended Data Table 4 | Mass spectrometry of sfGFP for identification and quantification of ncAA incorporation

Modified sequence	Modifications	Precursor m/z	Precursor charge	Retention time (min)	Mass error (ppm)	PEP	Score
FEGDTLVNR*	Unmodified	525.7644	2	28.86	0.18	0.00E+00	186.090
SAMPEGYVQER*	Unmodified	633.7928	2	31.73	–0.56	2.49E–03	18.312
SAM(ox)PEGYVQER*	Oxidation (M)	641.7903	2	20.61	–0.24	3.25E–07	60.102
LEYNFNSHNVNITADK	Unmodified	626.9709	3	39.28	–0.40	3.58E–09	36.193
LEYNFNSHNVCITADK	Unmodified	962.9465	2	42.92	1.46	1.22E–77	90.434
LEYNFNSHNVK†	Unmodified	682.8333	2	27.99	–0.46	1.01E–05	44.309
LEYNFNSHNVGITADK	Unmodified	607.9637	3	38.81	0.11	7.45E–03	16.306
LEYNFNSHNVN(pr)ITADK	PrK	987.9814	2	45.99	–0.01	0.00E+00	266.850
LEYN(de)FNSHNVN(pr)ITADK	Deamidation (N), PrK	659.3181	3	49.20	0.15	0.00E+00	128.900
LEYNFN(de)SHNVN(pr)ITADK	Deamidation (N), PrK	988.4734	2	49.13	–0.11	0.00E+00	121.660
LEYNFNSHN(de)VN(pr)ITADK	Deamidation (N), PrK	659.3181	3	49.42	0.31	0.00E+00	95.662
LEYN(de)FNSHNVSITADK	Deamidation (N)	926.9392	2	40.77	NaN	8.53E–280	60.628
LEYNFN(de)SHNVSITADK	Deamidation (N)	926.9392	2	41.25	–0.09	0.00E+00	127.710
LEYNFNSHN(de)VSITADK	Deamidation (N)	618.2952	3	41.32	0.62	8.93E–96	68.919
LEYNFNSHNVSITADK	Unmodified	926.4472	2	39.64	–0.26	0.00E+00	222.550
LEYNFNSHNVVITADK	Unmodified	621.9794	3	41.79	–0.27	7.95E–40	62.377
LEYNFNSHNVMITADK	Unmodified	632.6367	3	40.29	2.97	2.03E–02	10.057
LEYN(de)FNSHNVYITADK	Deamidation (N)	964.9549	2	46.10	0.52	2.44E–38	62.140
LEYNFN(de)SHNVYITADK	Deamidation (N)	643.6390	3	46.39	0.37	0.00E+00	91.457
LEYNFNSHN(de)VYITADK	Deamidation (N)	964.9549	2	46.39	–0.34	2.00E–39	56.951
LEYNFNSHNVYITADK	Unmodified	964.4629	2	43.24	–0.30	0.00E+00	309.980
LEYNFNSHNV(I/L)ITADK	Unmodified	626.6513	3	44.33	–0.24	7.78E–161	75.463

Modified sequence	Modifications	Precursor m/z	Precursor charge	Retention time (min)
SAMPEGYVQER*	Unmodified	633.7928	2	27.36
SAM(ox)PEGYVQER*	Oxidation (M)	641.7903	2	20.63
FEGDTLVNR*	Unmodified	525.7644	2	28.66
LEYNFNSHNVSITADK	Unmodified	617.9672	3	39.50
LEYNFNSHN(de)VSITADK	Deamidation (N)	618.2952	3	40.25
LEYNFNSHNVK†	Unmodified	455.5580	3	27.94
LEYNFNSHNVNITADK	Unmodified	626.9709	3	39.20
LEYNFNSHNVN(pr)ITADK	PrK	658.9900	3	45.71
LEYNFN(de)SHNVN(pr)ITADK	Deamidation (N), PrK	659.3181	3	49.19
LEYNFNSHNVCITADK	Unmodified	642.3001	3	45.47
LEYNFNSHNVGITADK	Unmodified	607.9637	3	38.81
LEYNFNSHNV(I/L)ITADK	Unmodified	626.6513	3	44.38
LEYNFNSHNVMITADK	Unmodified	632.6367	3	40.33
LEYNFNSHNVYITADK	Unmodified	643.3110	3	43.03
LEYNFN(de)SHNVYITADK	Deamidation (N)	643.6390	3	44.27
LEYNFNSHN(de)VYITADK	Deamidation (N)	643.6390	3	45.75
LEYNFNSHNVVITADK	Unmodified	621.9794	3	41.85
LEYN(de)FNSHNVN(pr)ITADK	Deamidation (N), PrK	659.3181	3	47.50

Summary of all reference and reporter peptides identified by database searching (upper table) and peptides used for precursor ion-based quantification (lower table). The highest scoring peptide for each identified peptide species is reported. PrK and natural amino acids at position 151 of sfGFP are denoted in red. Modifications are abbreviated as follows: (ox), oxidation of Met; N(pr), PrK; (de), deamidation of Asn. The isobaric amino acids Ile and Leu are indistinguishable in our experiments.
*sfGFP reference peptides used for normalization between samples; all other peptides are reporter peptides.
†Reporter peptide truncated owing to enzymatic cleavage by LysC and trypsin.

nature research

Corresponding author(s): Floyd E. Romesberg

☐ Initial submission ☐ Revised version ☒ Final submission

Life Sciences Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form is intended for publication with all accepted life science papers and provides structure for consistency and transparency in reporting. Every life science submission will use this form; some list items might not apply to an individual manuscript, but all fields must be completed for clarity.

For further information on the points included in this form, see Reporting Life Sciences Research. For further information on Nature Research policies, including our data availability policy, see Authors & Referees and the Editorial Policy Checklist.

Experimental design

1. Sample size

Describe how sample size was determined.

No sample size calculation was performed. Sample sizes were based on material constraints (i.e. the amount of unnatural triphosphates required to grow all the cultures used in the manuscript). As indicated in the text, all experiments were performed with n = 4 cultures (except one with n = 3) each inoculated from a single colony, which provides some limited information about the distribution of the measurements and is typical of similar experiments and studies in the field.

2. Data exclusions

Describe any data exclusions.

The plasmid cloning strategy described in the Methods results in a small percentage of plasmids that do not contain the UBP and/or contain other assembly errors during plasmid construction. Colonies transformed with such plasmids (as determined by the UBP retention assay described in the text) were excluded from analysis, as experiments conducted with cultures derived from such colonies are not relevant for assessing the ability of UBPs to be decoded in vivo. This exclusion criteria was pre-established in a previous study (see doi: 10.1073/pnas.1616443114).

3. Replication

Describe whether the experimental findings were reliably reproduced.

As indicated in the Methods, all experiments involving plated colonies were performed once due to material constraints. However, comparable experiments involving direct inoculation of electroporated cells into liquid media were also performed and all such experimental replicates were successful and reproduced the findings.

4. Randomization

Describe how samples/organisms/participants were allocated into experimental groups.

Bacterial colonies from transformations were selected randomly. Allocation into experimental groups is irrelevant; single colonies were propagated as bacterial cultures, which were then split and tested with various experimental conditions, where applicable.

5. Blinding

Describe whether the investigators were blinded to group allocation during data collection and/or analysis.

Blinding is not relevant to our study because none of our data is based on qualitative scoring metrics nor does it involve animals or human research participants. As described in the above section for Randomization, blinding during group allocation is irrelevant because the samples of bacterial cultures that were split into different conditions were random samplings and there is no control over which cells will be selected and thus, no bias during group allocation.

Note: all studies involving animals and/or human research participants must disclose whether blinding and randomization were used.

6. Statistical parameters

For all figures and tables that use statistical methods, confirm that the following items are present in relevant figure legends (or in the Methods section if additional space is needed).

n/a	Confirmed	
☐	☒	The exact sample size (*n*) for each experimental group/condition, given as a discrete number and unit of measurement (animals, litters, cultures, etc.)
☐	☒	A description of how samples were collected, noting whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
☐	☒	A statement indicating how many times each experiment was replicated
☒	☐	The statistical test(s) used and whether they are one- or two-sided (note: only common tests should be described solely by name; more complex techniques should be described in the Methods section)
☒	☐	A description of any assumptions or corrections, such as an adjustment for multiple comparisons
☒	☐	The test results (e.g. *P* values) given as exact values whenever possible and with confidence intervals noted
☐	☒	A clear description of statistics including central tendency (e.g. median, mean) and variation (e.g. standard deviation, interquartile range)
☒	☐	Clearly defined error bars

See the web collection on statistics for biologists for further resources and guidance.

Software

Policy information about availability of computer code

7. Software

Describe the software used to analyze the data in this study.

Basic statistical analysis was performed using Microsoft Excel. Chemiluminescent western blots were processed in UVP VisionWorks® LS (v 8.6.15114.8618) using the noise removal setting (Starfield Subtraction) and then further processed (black-white inverted) using Image Studio Lite (v. 5.2.5, LI-COR Biosciences). Fluorescent western blots were processed (pseudo-colored) and quantified using Image Studio Lite (v. 5.2.5, LI-COR Biosciences). Fluorescence overlays between the anti-GFP western blots and TAMRA fluorescence scans were performed with Paint.NET (v. 3.5.10). Biotin shift gels were quantified using Image Studio Lite (v. 5.2.5, LI-COR Biosciences). Peptide database searches were performed with MaxQuant (v. 1.5.8.0) using the default search parameters with minor changes, as described in Methods. Quantitation of LC/MS chromatographic peak intensities was performed by extracting MS1 precursor ion chromatograms with Skyline Software (v. 3.6.0.10167).

For manuscripts utilizing custom algorithms or software that are central to the paper but not yet described in the published literature, software must be made available to editors and reviewers upon request. We strongly encourage code deposition in a community repository (e.g. GitHub). *Nature Methods* guidance for providing algorithms and software for publication provides further information on this topic.

Materials and reagents

Policy information about availability of materials

8. Materials availability

Indicate whether there are restrictions on availability of unique materials or if these materials are only available for distribution by a for-profit company.

As indicated in the Methods, the unnatural nucleotide triphosphates are available by custom chemical synthesis from commercial vendors (WuXi AppTec, Shanghai, China for nucleosides and TriLink BioTechnologies LLC, San Diego, USA or MyChem LLC, San Diego, USA for triphosphorylation of the nucleosides)

9. Antibodies

Describe the antibodies used and how they were validated for use in the system under study (i.e. assay and species).

Rabbit anti-GFP antibody (product #G1544, lot 046M4871V, Sigma-Aldrich)
Goat anti-rabbit-Alexa Fluor 647-conjugated antibody (product #A32733, lot #SD250298, ThermoFisher)
Goat anti-rabbit-HRP (product #170-6515, lot #64106148, BioRad)
All antibodies are validated for the application used in this study (Western blotting) according to the respective manufacturers' websites

10. Eukaryotic cell lines

a. State the source of each eukaryotic cell line used. n/a

b. Describe the method of cell line authentication used. n/a

c. Report whether the cell lines were tested for mycoplasma contamination. n/a

d. If any of the cell lines used are listed in the database of commonly misidentified cell lines maintained by ICLAC, provide a scientific rationale for their use. n/a

Animals and human research participants

Policy information about studies involving animals; when reporting animal research, follow the ARRIVE guidelines

11. Description of research animals

Provide details on animals and/or animal-derived materials used in the study. n/a

Policy information about studies involving human research participants

12. Description of human research participants

Describe the covariate-relevant population characteristics of the human research participants. n/a